

# Soil improvers and growing media — Extraction of calcium chloride/DTPA (CAT) soluble nutrients

The European Standard EN 13651:2001 has the status of a  
British Standard

ICS 65.080

# National foreword

This British Standard is the official English language version of EN 13651:2001.

The UK participation in its preparation was entrusted to Technical Committee AW/20, Top soil and other growing media, which has the responsibility to:

- aid enquirers to understand the text;
- present to the responsible European committee any enquiries on the interpretation, or proposals for change, and keep the UK interests informed;
- monitor related international and European developments and promulgate them in the UK.

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This British Standard, having been prepared under the direction of the Consumer Products and Services Sector Policy and Strategy Committee, was published under the authority of the Standards Policy and Strategy Committee on 09 November 2001

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English version

## Soil improvers and growing media - Extraction of calcium chloride/DTPA (CAT) soluble nutrients

Amendements du sol et supports de culture - Extraction  
des éléments nutritifs solubles dans le chlorure de  
calcium/DTPA (CAT)

Bodenverbesserungsmittel und Kultursubstrate - Extraktion  
von in Calciumchlorid/DTPA (CAT) löslichen Nährstoffen

This European Standard was approved by CEN on 11 August 2001.

CEN members are bound to comply with the CEN/CENELEC Internal Regulations which stipulate the conditions for giving this European Standard the status of a national standard without any alteration. Up-to-date lists and bibliographical references concerning such national standards may be obtained on application to the Management Centre or to any CEN member.

This European Standard exists in three official versions (English, French, German). A version in any other language made by translation under the responsibility of a CEN member into its own language and notified to the Management Centre has the same status as the official versions.

CEN members are the national standards bodies of Austria, Belgium, Czech Republic, Denmark, Finland, France, Germany, Greece, Iceland, Ireland, Italy, Luxembourg, Netherlands, Norway, Portugal, Spain, Sweden, Switzerland and United Kingdom.



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## Contents

	page
Foreword .....	3
1 Scope.....	4
2 Normative references.....	4
3 Terms and definitions.....	4
4 Principle .....	4
5 Reagents .....	4
6 Apparatus.....	5
7 Test sample.....	5
8 Procedure.....	6
9 Determination of the extracted nutrients and elements.....	6
10 Expression of results.....	6
11 Precision .....	6
12 Test report.....	7
Annex A (informative) Results of an interlaboratory trial to determine calcium chloride/DTPA (CAT) soluble nutrients and elements.....	8
Annex B (informative) Methods of analysis used in the interlaboratory trial.....	15
Bibliography.....	16

## Foreword

This European Standard has been prepared by Technical Committee CEN/TC 223 "Soil improvers and growing media", the secretariat of which is held by BSI.

This European Standard shall be given the status of a national standard, either by publication of an identical text or by endorsement, at the latest by March 2002, and conflicting national standards shall be withdrawn at the latest by March 2002.

The annexes A and B are informative.

According to the CEN/CENELEC Internal Regulations, the national standards organizations of the following countries are bound to implement this European Standard: Austria, Belgium, Czech Republic, Denmark, Finland, France, Germany, Greece, Iceland, Ireland, Italy, Luxembourg, Netherlands, Norway, Portugal, Spain, Sweden, Switzerland and the United Kingdom.



**SAFETY PRECAUTIONS** — Care should be taken when handling samples that may contain sharps or is of a dusty nature.

## 1 Scope

This European Standard specifies an extraction method for the routine determination of calcium chloride/DTPA (CAT-method) extractable nutrients and elements (as listed in annex B) in soil improvers or growing media.

The method is not applicable to liming materials and preformed materials such as mineral wool slabs and foam slabs.

**NOTE** The requirements of the standard may differ from the national legal requirements for the declaration of the products concerned.

## 2 Normative references

This European Standard incorporates by dated or undated reference, provisions from other publications. These normative references are cited at the appropriate places in the text, and the publications are listed hereafter. For dated references, subsequent amendments to or revisions of any of these publications apply to this European Standard only when incorporated in it by amendment or revision. For undated references the latest edition of the publication referred to applies (including amendments).

EN ISO 3696, Water for analytical laboratory use - Specification and test methods (ISO 3696:1987).

EN 13040:1999, Soil improvers and growing media - Sample preparation for chemical and physical tests, determination of dry matter content, moisture content and laboratory compacted bulk density.

## 3 Terms and definitions

For the purposes of this standard the terms and definitions given in EN 13040 apply.

## 4 Principle

A sample is extracted with calcium chloride/DTPA at 22 C ± 3 C in an extraction volume ratio of 1+5. The extracted nutrients are determined by various methods as appropriate (see annex B).

## 5 Reagents

### 5.1 General

All reagents used shall be of recognised analytical quality and water of grade 2 according to EN ISO 3696.

**5.2 Dihydrated calcium chloride,  $\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$ .**

**5.3 Diethylenetriaminepentaacetic acid (DTPA),  $\text{C}_{14}\text{H}_{23}\text{N}_3\text{O}_{10}$ .**

**5.4 Concentrated extracting solution  $\text{CaCl}_2$ /DTPA, (CAT),** dissolve 14,7 g  $\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$  (5.2) and 7,88 g DTPA (5.3) in 800 ml hot water (temperature approximately 80 C) with stirring on a magnetic stirrer in a 1000 ml beaker. At 75 C ± 10 C the reagents will dissolve within 2 h. Allow to cool to ambient temperature. Transfer the solution to a 1000 ml flask and dilute to the mark with water. The

solution is stable at room temperature for several weeks. Any precipitation that occurs will disappear with warming and stirring.

**5.5 Extracting solution  $\text{CaCl}_2/\text{DTPA}$ , (CAT),** dilute the concentrated extracting solution (5.4) with water in the proportions one part concentrated solution with nine parts water. The final concentration of the extracting solution should be 0,01 mol/l  $\text{CaCl}_2$  and 0,002 mol/l DTPA. The pH of the extracting solution should be adjusted if necessary to be between 2,6 and 2,65.

**5.6 Nitric acid,**  $c(\text{HNO}_3) = 15 \text{ mol/l}$ : 1,42 g/ml; not less than 65 % mass/volume.

**5.7 Nitric acid,**  $c(\text{HNO}_3) = 0,5 \text{ mol/l}$ , dilute 35 ml nitric acid (5.6) to one litre with water.

## 6 Apparatus

### 6.1 General

**NOTE** It has been found convenient to keep separate sets of glassware for the determinations given in annex B, in order to reduce the possibility of within-laboratory contamination.

All glass apparatus and plastic vessels used in the procedure should be subject to an appropriate preparation procedure in order to keep the risk of contamination as low as possible. It is recommended that all vessels (glass and plastic) are cleansed by carefully immersing in warm nitric acid (5.6) for a minimum of 6 h and then rinsed in water.

Rubber stoppers, which may contain traces of metals shall not be used. It is recommended to use plastic or any other stopper free of all substances to be analyzed.

Borosilicate glassware shall not be used if boron is to be determined.

The apparatus consists of the usual laboratory apparatus, and in particular the following :

**6.2 Plastic bottles or containers,** sufficiently large (500 ml to 1500 ml) with screw cap to accommodate the volume of the sample, extractant and at least 10 % air volume.

**6.3 Shaking or mixing machine,** capable of holding the plastic bottles or containers (6.2) and maintaining the sample in suspension without damaging the structure of the sample. The use of a horizontal table shaker is recommended.

**6.4 Filter paper,** cellulose-based ashless types, with a medium pore size of approximately 8  $\mu\text{m}$  and diameter of 150 mm.

**NOTE** Centrifugation is an acceptable alternative.

**6.5 Analytical balance** with an accuracy of 10 mg.

## 7 Test sample

Prepare the laboratory sample in accordance with EN 13040:1999, clause 8, and determine the laboratory compacted bulk density of the sample in accordance with EN 13040:1999, annex A.

## 8 Procedure

### 8.1 Extraction

#### 8.1.1 Test samples passing through a 20 mm sieve

Take a weight equivalent to 60 ml of the sample volume (EN 13040:1999, clause 8.5) to the nearest 1 g and transfer to the container (6.2). Add 300 ml of extracting solution (5.5), secure the cap and shake for 1 h on the shaking machine (6.3) at 22 C  $\pm$  3 C.

#### 8.1.2 Test samples passing through a 40 mm sieve

Take a weight equivalent to 250 ml of the sample volume (EN 13040:1999, clause 8.3) to the nearest 1 g and transfer to the container (6.2). Add 1250 ml of extracting solution (5.5), secure the cap and shake for 1 h on the shaking machine (6.3) at 22 C  $\pm$  3 C.

#### 8.1.3 Filtration

Filter through filter paper (6.4) discarding at least the first 10 ml. In some cases paper filtration is too slow or even impossible. In such cases alternative procedures for obtaining a clear supernatant are acceptable and the technique used shall be reported. The filtered extract is stable for three days in a hermetically closed polyethylene bottle if stored in a refrigerator at 0 C to 5 C. The filtrate can be stored for longer periods in a deep freezer at about  $-18$  C.

**NOTE** Before using a solution that has been frozen, the thawed solution shall be thoroughly mixed to eliminate gradient separation that occurs on freezing and subsequent thawing.

### 8.2 Blank

The reagent blank test shall be carried out in parallel with the determination, by the same procedure as outlined in 8.1.1 or 8.1.2 and 8.1.3, using the same quantities of all the reagents as in the determination but omitting the test portion.

**NOTE** The measurement of a blank is introduced to determine the contribution of the extracting solution, glassware and filter paper used.

## 9 Determination of the extracted nutrients and elements

See annex B.

## 10 Expression of results

Subtract the values determined for the reagent blank from those obtained for the samples. All results shall be calculated using the determined compacted laboratory bulk density and expressed in mg/l substrate as received basis.

## 11 Precision

The repeatability and reproducibility of the calcium chloride DTPA soluble nutrient content in separately prepared samples should be in accordance with Tables A.1 to A.6.

A summary of the results of an interlaboratory trial to determine the precision of the method, in accordance with ISO 5725 [2], is given in annex A.

**NOTE** The values derived from this interlaboratory trial may not be applicable to concentrations and matrices other than those tested.



## 12 Test report

The test report shall contain the following information:

- a) a reference to this European Standard;
- b) all information necessary for complete identification of the sample;
- c) all the analytical methods used;
- d) the results of the determination, expressed as mg/l fresh sample;
- e) details of any operations not specified in the European Standard or regarded as optional, as well as any factor which may have affected the results;
- f) the laboratory compacted bulk density result.

## **Annex A** (informative)

### **Results of an interlaboratory trial to determine calcium chloride/DTPA (CAT) soluble nutrients and elements**

An interlaboratory trial was organized in 1997 under the auspices of the European Committee for Standardization, to test the procedures specified in this European Standard.

In this trial the number of laboratories given in the Tables A.1 to A.6 determined the calcium chloride/DTPA soluble nutrient and elements content in six sample types.

All results are reported on a fresh basis.

A summary of the results of the interlaboratory trial for the determination of calcium chloride/DTPA (CAT) soluble nutrients and elements in six sample types is given in Tables A.1 to A.6.

Table A.1 - Composted bark

Element	No. of labs after eliminating outliers	No. of outliers (labs)	Mean value mg/l	Repeatability $s_r$	Repeatability Std Dev %	Repeatability limit $r = 2,8 s_r$	Reproducibility $s_R$	Reproducibility Std Dev %	Reproducibility limit $R = 2,8 s_R$
NH <sub>4</sub> -N	19	0	1220,1	49,19	11,29	137,72	248,52	57,03	695,87
NO <sub>3</sub> -N	13	2	13,6	2,27	46,65	6,35	13,96	287,18	39,08
P	20	0	290,0	14,59	14,09	40,85	108,99	105,22	305,17
K	21	0	2727,0	71,12	7,30	199,13	401,48	41,22	1124,15
Mg	20	1	170,4	7,33	12,05	20,52	57,22	94,04	160,21
SO <sub>4</sub> -S	13	0	100,7	8,93	24,82	24,99	47,53	132,15	133,09
Na	19	0	389,7	7,83	5,63	21,93	59,44	42,71	166,42
B	13	0	1,8	0,09	14,14	0,25	0,74	117,42	2,06
Cu	18	1	1,1	0,08	20,68	0,23	0,25	63,39	0,70
Fe	17	2	14,0	0,92	18,48	2,58	2,31	46,28	6,47
Mn	18	1	10,7	0,48	12,72	1,36	1,78	46,79	4,99
Mo	9	1	0,1	0,02	43,46	0,06	0,04	88,68	0,11
Zn	18	1	15,3	0,69	12,60	1,93	2,94	53,79	8,23
Cd	7	1	0,02	0,00	46,84	0,01	0,01	182,32	0,04
Pb	9	1	0,25	0,03	35,29	0,09	0,17	188,29	0,47
Element	No. of labs after eliminating outliers	No. of outliers (labs)	Mean value g/l	Repeatability $s_r$	Repeatability Std Dev %	Repeatability limit $r = 2,8 s_r$	Reproducibility $s_R$	Reproducibility Std Dev %	Reproducibility limit $R = 2,8 s_R$
CLBD <sup>a</sup>	14	1	392,8	8,3	5,9	23,3	22,1	15,8	61,9
<sup>a</sup> CLBD - Compacted laboratory bulk density									

Table A.2 - Biowaste

Element	No. of labs after eliminating outliers	No. of outliers (labs)	Mean value mg/l	Repeatability $s_r$	Repeatability Std Dev %	Repeatability limit $r = 2,8 s_r$	Reproducibility $s_R$	Reproducibility Std Dev %	Reproducibility limit $R = 2,8 s_R$
NH <sub>4</sub> -N	19	0	192,0	9,87	14,39	27,63	79,02	115,22	221,25
NO <sub>3</sub> -N	16	3	33,5	1,35	11,25	3,77	27,34	228,58	76,56
P	18	2	42,9	1,30	8,48	3,64	12,54	81,85	35,11
K	21	1	2240,8	46,61	5,82	130,52	446,10	55,74	1249,08
Mg	21	1	192,3	5,32	7,75	14,90	39,65	57,75	111,02
SO <sub>4</sub> -S	13	2	52,5	0,74	3,95	2,08	12,48	66,54	34,94
Na	19	0	210,1	8,72	11,62	24,42	37,02	49,33	103,66
B	12	0	2,5	0,12	14,15	0,35	0,79	90,13	2,21
Cu	19	0	1,6	0,10	17,34	0,28	0,20	34,33	0,55
Fe	19	4	59,4	3,13	14,73	8,75	7,12	33,53	19,93
Mn	16	3	36,0	1,47	11,42	4,11	6,79	52,78	19,01
Mo	9	0	0,1	0,01	35,77	0,03	0,04	105,22	0,10
Zn	17	2	20,3	1,10	15,23	3,09	1,84	25,34	5,15
Cd	7	1	0,1	0,00	7,29	0,00	0,03	160,66	0,09
Pb	12	0	7,4	0,45	17,10	1,26	1,18	44,78	3,29
Ni	8	1	0,2	0,06	86,56	0,16	0,11	169,67	0,31
Element	No. of labs after eliminating outliers	No. of outliers (labs)	Mean value g/l	Repeatability $s_r$	Repeatability Std Dev %	Repeatability limit $r = 2,8 s_r$	Reproducibility $s_R$	Reproducibility Std Dev %	Reproducibility limit $R = 2,8 s_R$
CLBD <sup>a</sup>	15	1	649,5	3,5	1,5	9,7	22,7	9,8	63,6
<sup>a</sup> CLBD - Compacted laboratory bulk density									

Table A.3 - Clay peat (fertilized)

Element	No. of labs after eliminating outliers	No. of outliers (labs)	mean value mg/l	Repeatability  $s_r$	Repeatability Std Dev %	Repeatability limit $r = 2,8 s_r$	Reproducibility  $s_R$	Reproducibility Std Dev %	Reproducibility limit $R = 2,8 s_R$
NH <sub>4</sub> -N	19	0	87,2	2,06	6,63	5,78	11,81	37,93	33,08
NO <sub>3</sub> -N	19	0	72,9	1,77	6,81	4,97	11,89	45,67	33,31
P	19	1	35,2	1,19	9,45	3,32	4,62	36,73	12,93
K	20	1	148,0	3,02	5,72	8,46	18,91	35,78	52,94
Mg	19	2	84,9	1,30	4,28	3,63	6,46	21,31	18,09
SO <sub>4</sub> -S	13	0	107,5	4,02	10,48	11,26	12,57	32,76	35,20
Na	18	0	28,5	1,15	11,28	3,21	5,35	52,54	14,97
B	8	2	0,1	0,01	12,52	0,02	0,08	151,80	0,22
Cu	19	0	0,7	0,04	17,93	0,12	0,17	69,54	0,48
Fe	17	2	46,8	1,43	8,55	4,00	6,81	40,79	19,08
Mn	19	0	9,8	0,27	7,65	0,75	1,47	42,07	4,12
Mo	4	2	0,1	0,00	12,21	0,01	0,01	48,86	0,03
Zn	18	1	1,4	0,04	7,96	0,11	0,20	40,52	0,56
Cd	9	0	0,0	0,01	65,87	0,02	0,01	69,03	0,02
Pb	12	0	0,9	0,05	15,64	0,15	0,12	36,72	0,34
Ni	9	0	0,1	0,02	37,67	0,04	0,04	103,40	0,12
Element	No. of labs after eliminating outliers	No. of outliers (labs)	Mean value g/l	Repeatability  $s_r$	Repeatability Std Dev %	Repeatability limit $r = 2,8 s_r$	Reproducibility  $s_R$	Reproducibility Std Dev %	Reproducibility limit $R = 2,8 s_R$
CLBD <sup>a</sup>	16	0	349,2	11,0	8,8	30,8	35,5	28,5	99,4
<sup>a</sup> CLBD - Compacted laboratory bulk density									



Table A.4 - Coarse peat (fertilized)

Element	No. of labs after eliminating outliers	No. of outliers (labs)	Mean value mg/l	Repeatability  s <sub>r</sub>	Repeatability Std Dev %	Repeatability limit r = 2,8 s <sub>r</sub>	Reproducibility  s <sub>R</sub>	Reproducibility Std Dev %	Reproducibility limit R = 2,8 s <sub>R</sub>
NH <sub>4</sub> -N	20	0	105,1	5,18	13,80	14,50	15,86	42,28	44,42
NO <sub>3</sub> -N	20	0	53,6	4,22	22,06	11,82	8,94	46,72	25,03
P	21	0	92,9	9,41	28,36	26,34	25,12	75,72	70,33
K	20	2	129,3	8,25	17,87	23,11	22,92	49,62	64,16
Mg	21	1	251,0	17,50	19,52	49,00	38,31	42,75	107,28
SO <sub>4</sub> -S	14	0	107,1	10,88	28,44	30,47	20,55	53,72	57,55
Na	19	1	35,3	1,52	12,10	4,27	6,30	50,00	17,63
B	13	0	0,2	0,04	45,35	0,11	0,08	90,57	0,21
Cu	19	1	1,1	0,11	27,24	0,30	0,21	53,71	0,59
Fe	20	0	9,9	0,87	24,57	2,44	1,96	55,21	5,48
Mn	20	0	3,3	0,29	25,16	0,83	0,79	67,50	2,21
Mo	10	0	0,5	0,11	65,52	0,31	0,11	67,18	0,32
Zn	17	2	0,7	0,12	45,67	0,34	0,18	66,73	0,50
Cd	9	0	0,02	0,00	22,79	0,00	0,00	70,15	0,01
Pb	10	0	0,2	0,05	75,60	0,15	0,07	106,83	0,21
Element	No. of labs After eliminating outliers	No. of outliers (labs)	Mean value g/l	Repeatability  s <sub>r</sub>	Repeatability Std Dev %	Repeatability limit r = 2,8 s <sub>r</sub>	Reproducibility  s <sub>R</sub>	Reproducibility Std Dev %	Reproducibility limit R = 2,8 s <sub>R</sub>
CLBD <sup>a</sup>	16	0	224,8	7,6	9,5	21,4	19,5	24,3	54,6
<sup>a</sup> CLBD - Compacted laboratory bulk density									

Table A.5 - Composted sludge

Element	No. of labs after eliminating outliers	No. of outliers (labs)	Mean value mg/l	Repeatability $s_r$	Repeatability Std Dev %	Repeatability limit $r = 2,8 s_r$	Reproducibility $s_R$	Reproducibility Std Dev %	Reproducibility limit $R = 2,8 s_R$
NH <sub>4</sub> -N	18	1	521,7	14,82	7,96	41,50	169,81	91,14	475,48
NO <sub>3</sub> -N	19	0	243,5	9,72	11,18	27,21	83,94	96,54	235,04
P	19	0	132,6	3,85	8,13	10,77	52,11	110,05	145,90
K	21	0	2584,3	48,36	5,24	135,40	406,49	44,04	1138,17
Mg	18	1	223,5	7,47	9,36	20,93	50,45	63,21	141,27
SO <sub>4</sub> -S	13	0	199,8	7,79	10,92	21,82	30,35	42,54	84,98
Na	19	0	240,2	4,29	5,00	12,02	28,42	33,13	79,59
B	11	2	3,1	0,11	9,99	0,30	0,46	42,16	1,29
Cu	18	1	1,7	0,09	15,20	0,26	0,40	65,39	1,11
Fe	16	3	52,5	1,83	9,77	5,13	10,46	55,80	29,28
Mn	18	1	28,0	2,06	20,57	5,76	6,86	68,60	19,21
Mo	8	0	0,1	0,02	44,09	0,05	0,06	147,53	0,17
Zn	18	1	20,7	0,82	11,16	2,31	3,91	53,00	10,96
Cd	7	2	0,1	0,01	42,17	0,03	0,04	141,29	0,12
Pb	12	0	3,5	0,16	13,10	0,46	0,51	40,95	1,43
Element	No. of labs after eliminating outliers	No. of outliers (labs)	Mean value g/l	Repeatability $s_r$	Repeatability Std Dev %	Repeatability limit $r = 2,8 s_r$	Reproducibility $s_R$	Reproducibility Std Dev %	Reproducibility limit $R = 2,8 s_R$
CLBD <sup>a</sup>	14	1	535,5	12,7	6,6	35,4	26,1	13,6	73,0

<sup>a</sup> CLBD - Compacted laboratory bulk density

Table A.6 - Composted wood fibre

Element	No. of labs after eliminating outliers	No. of outliers (labs)	Mean value mg/l	Repeatability s <sub>r</sub>	Repeatability Std Dev %	Repeatability limit r = 2,8 s <sub>r</sub>	Reproducibility s <sub>R</sub>	Reproducibility Std Dev %	Reproducibility limit R = 2,8 s <sub>R</sub>
NH <sub>4</sub> -N	20	0	71,1	2,25	8,85	6,29	23,15	91,21	64,83
NO <sub>3</sub> -N	19	1	65,4	2,23	9,53	6,23	7,05	30,18	19,73
P	20	1	55,2	1,62	8,20	4,53	11,20	56,82	31,37
K	20	2	152,3	4,33	7,96	12,11	14,77	27,15	41,35
Mg	22	0	71,7	2,77	10,80	7,75	8,13	31,72	22,75
SO <sub>4</sub> -S	13	1	86,5	3,68	11,92	10,31	12,60	40,78	35,27
Na	19	1	23,3	1,08	13,01	3,02	3,41	41,03	9,54
B	12	0	0,2	0,03	44,04	0,09	0,09	122,30	0,24
Cu	20	0	1,0	0,04	12,36	0,12	0,18	51,00	0,49
Fe	18	2	38,0	0,98	7,22	2,74	4,86	35,78	13,60
Mn	20	0	8,5	0,36	11,91	1,01	0,85	28,03	2,39
Mo	8	0	0,2	0,02	32,75	0,05	0,05	87,81	0,14
Zn	20	0	1,6	0,06	10,70	0,17	0,20	36,23	0,57
Cd	10	0	0,0	0,00	38,18	0,01	0,00	55,02	0,01
Pb	11	1	0,8	0,03	11,59	0,10	0,13	43,83	0,37
Element	No. of labs after eliminating outliers	No. of outliers (labs)	Mean value g/l	Repeatability s <sub>r</sub>	Repeatability Std Dev %	Repeatability limit r = 2,8 s <sub>r</sub>	Reproducibility s <sub>R</sub>	Reproducibility Std Dev %	Reproducibility limit R = 2,8 s <sub>R</sub>
CLBD <sup>a</sup>	15	0	263,5	13,5	14,3	37,7	30,1	32,0	84,3
<sup>a</sup> CLBD - Compacted Laboratory Bulk Density									

## Annex B (informative)

### Methods of analysis used in the interlaboratory trial

<b><u>ELEMENT</u></b>	<b><u>METHOD</u></b>
Ammonium-N	4, 6, 7, 8,10
Nitrate-N	4, 6, 8, 9, 10
Phosphorus	1, 4, 11
Potassium	1, 2, 3
Magnesium	1, 2
Sulfate S	1, 4
Sodium	1, 2, 3
Boron	1, 5, 13
Copper	1, 2
Iron	1, 2
Manganese	1, 2
Molybdenum	1, 2, 12
Zinc	1, 2
Cadmium	1, 2
Nickel	1, 2
Lead	1, 2

#### Method

1	ISO 11885: 1996 Inductively coupled plasma – atomic emission spectrometry [10]
2	ISO 11047: 1998 Flame or furnace atomic absorption spectrometry [8]
3	ISO 9964-3:1993 Flame emission spectrometry [6]
4	ISO 10304-2: 1995 Ion chromatography [7]
5	Hoffmann 1997 Dianthrimide method [11]
6	ISO 5664: 1984 Distillation [1]
7	ISO 7150-1: 1984 Manual spectrophotometric [3]
8	ISO 11732: 1997 Flow spectrophotometric [9]
9	ISO 7890-3: 1988 Spectrometric - sulphosalic acid [5]
10	ISO 7150-2: 1986 Automated spectrophotometric method [4]
11	Hoffmann 1966 Ammonium molybdate ascorbic acid/stannous chloride reduction [12]
12	Reference Book 427 1986 spectrophotometric -iron/thiocyanate [13]
13	John M.K. et al., 1975, azomethine-H [14]

Alternative methods may be suitable for the concentration range and extract used. The user is to confirm that the method chosen gives results equivalent to those obtained by the methods listed above.

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